

SCREENING OF MYCOFLORA ASSOCIATED WITH MARKET DISEASES OF BANANA (MUSA PARADISICA)



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Fruits are best source of carbohydrates, antioxidants like vit-C, vit-E, vit- A & zinc, selenium, lycopene etc. There are reports from different parts of world that antioxidants play important role in antiaging process. Human body requires different types of essential amino acids all such amino acid are present in fruits. Amino acids prevent infections of pathogens.

Fruits improve overall health of human beings. Strong immune system increases resistance power towards different types of diseases. Fruits provide all types of minerals that are required to our body. The nutritionists placed fruits and green vegetables on the top because; fruits contain all types of essential ingredients which are required for healthy living being. Fruits contain 50-90% moisture.

The loss of fruits after harvest is a major problem of fruit growing countries of the world. There are reports of losses of millions of rupees annually due to post-harvest diseases caused by different types of mycoflora during transportation and storage periods. In order to make

plan for the disease control, it is very essential to have knowledge of pathogenic organism associated with fruits during storage periods. The associated mycoflora responsible for fruit losses. The aim of study of fruit diseases of market is to develop perfect disease management strategies that are economically significant. The diseases causes change in color, shape and biochemical alteration of the fruit due to interaction of the pathogen and the fruit becomes unfit for consumption.

The fruits are highly sensitive to the exogenous agencies such as fungi, bacteria etc. that affects the quality of fruit and causes loss of fruit. The diseases of fruits which occur after harvest during packing, transportation and storage periods due to fungal or bacterial pathogen are called as post-harvest diseases, Sohi, H.S. (1975).

There are different types of fungi associated with market diseases of banana. Cherian, T.T., Rhizopus sp. from post-harvest rot of guava caused due to Rhizopus sp. (2005) studied from. Sumia, et. al, (2006) reported white rot of papaya by caused by Sclerotium rolfsii. Cherian, T.T., (2005) studied post harvest rot of apple disease during storage which was due to infection of *Phytopthora colocaceae*. Fusarium rot of Peach (Choudhary, (1950).); Alternaria and Cladosporium rot of grapes fruits (Ali, 2010). Rhizopus on grapes (Rathod, G. M., 2010). In Marathwada region fruit storage system is not well developed. There is no any facility for storage of fruits after harvest from government. The roads are not well. Hence during transportation many of the fruits get injured. The injured fruits immediately get attacked by saprophytic fungi which causes rot diseases during transportation or storage period. Because of such problems about 20-55 percent fruits get deteriorated due to mycoflora. Mostly the fruits are stored by traditional method which has adverse effect on health of stored fruits. The total estimated loss comes to lakhs of rupees annually. Some of the pathogenic fungi attack fruits at immature stage and some others infect to overripe fruits. Major loss of harvested fruits is caused due to fungi. The diseases caused before harvest of fruits also responsible for degradation of quality and cost of banana in market.

Material and Method: The banana fruits were collected from different fruit store houses of different go downs of Aurangabad. The infected fruits of each type infection collected in separate from the same lot 10 immature healthy fruits were collected in a sterile polyethylene bag and from the same lot 10 immature healthy banana brought to the laboratory in another polythin bag. A separate polyethylene bag was used for each fruit. One infected fruit in one bag (Sumia Fatima, et. al., (2006). A separate bag was used even for healthy immature fruits (Bagwan, 2010). The mycoflora responsible for storage rot disease of banana fruit was isolated on PDA (Potato Dextrose Agar) medium by food poisoning technique method.

Before inoculation the infected fruit was surface sterilized with the help of 0.1% HgCl2 solution and then rinsed with the solution of sterile distilled water for 3-5 times to remove traces of HgCl2 solution. Then a small piece of infected region of fruit was removed with the help of sterile r needle and the infected portion was inoculated on PDA (Potato Dextrose Agar) medium amended petriplate in sterile condition. The inoculated petriplate were incubated at room temperature 24+-^O C. The fungus growing from the inoculated infected piece was inoculated on PDA medium. To get pure culture of the fungus a single hypha was removed from the inoculated petriplate and inoculated on freshly prepared PDA medium amended petriplate. To find out the Pathogenecity of the isolated fungus, a 4mm disc of growing colony was removed by sterile borer in sterile condition and inoculated on respective fruit. A set of 5 fruits was used to confirm pathogenecity. The pathogenecity was confirmed by following Koch's postulates. The fungi were identified on the basis of morphological features, type of colony growth, colour of colony, size and shape of spores and pigmentation.

Table: Post harvest mycoflora associated with diseases of Banana (Musa paradisica)

1.	Banana (//	Musa	Rhizopus oryzae; Macrophomina phaseolina; Colletotrichum
	paradisica)		musae; Chetothyrina musarum ;Fusarium moniliforme; Phoma
			jolyana; Fusarium roseum; Alternaria alternata; Verticillium
			albo- atrum; Ceratocystis paradoxa; Colletotrichum musae;
			Botryodiploidia theobromae; Trichothecium roseum;
			Macrophoma musae; Phytopthora nicotianae; Verticillium
			theobromae; Helminthosporium torulosum; Nigrospora oryzae;
			Pestalotia leprilegna ;Cladosporium; Rhizopus stolonifer;
			Gloeosporium musarum; Phoma herbarum; Fusarium
			oxysporum; Curvularia lunata; Phytopthora palmivora.

Result: The table shows the list of mycoflora causing market diseases of fungi isolated from banana (*Musa paradisiaca* L.). There were about 26 types of fungi isolated from infected fruits of banana. During collection of fruit samples from various parts go downs of Aurangabad. It was observed that there was numerous types of symptoms were observed due to infection and growth of post-harvest fungi. It is noted that some of the fungal genera showing a broad spectrum of host range. The forms of mycoflora like *Aspergillus*, *Alternaria*, *Curvularia*, *Fusarium*, *Mucor*, *Rhizopus*, *Colletotrichum*, *Penicillium*, *Phytopthora* and *Pestalotia* were found on 20 samples of fruits collected from different go downs. Among the fungal isolates the genus *Aspergillus*, *Rhizopus* and *Fusarium* was found more destructive

and was associated with all rotted fruits collected from various god owns. The post-harvest diseases of fruits were studied by Droby, et. al, (2002) isolated six types of fungi from infected banana fruits. By avoiding contact of fruits with field soil during harvest can reduce infection of most of the fungi causing post-harvest diseases of fruits to avoid economic losses. This practice can reduce disease intensity during storage periods. Proper handling of fruits during harvest, careful grading, proper selection of packing material, are very important factors to avoid infection of post-harvest fungi.

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